



CASE REPORT

De novo variant in the COL1A1 gene associated with orphan genetic disease: Osteogenesis Imperfecta type I

Jhonatan Alzate Valencia^{1,2,3} , Lina-Johanna Moreno-Giraldo^{1,2,3} 

1) Clinical Postgraduate Program in Pediatrics, Faculty of Health Sciences, Universidad Libre Cali, Colombia;

2) Research Group in Pediatrics (GRINPED); 3) Research Line in Neurogenetics and Metabolic Diseases.

Received: May 4th 2024 / Accepted: July 28th 2024 / Published: November 20th, 2024

© The Author(s) 2024. Article published with Open Access.



Abstract

Introduction: Osteogenesis Imperfecta (OI), is known as a disorder characterized by bone weakness and high risk of fractures, of genetic origin, known as brittle bone disease. It has a high burden of morbidity and mortality, associated with the presence of pain and functional limitation for those who present it. Fragility in the bones has been described by structural variants of the Collagen 1 genes (COL1A1 and COL1A2) among others, with an estimated prevalence between 1/15,000 and 1/20,000 cases in live newborns. The timely diagnosis of the disease allows to initiate targeted therapeutic alternatives, prevent complications, provide genetic counseling on risk of heritability, follow-up guidelines, prognosis approaching personalized medicine. **Case Presentation:** Female patient 8 years old, with clinical history of several fractures secondary to mild trauma (metatarsal left foot, right hand finger and ipsilateral foot), with other diagnoses of bilateral otosclerosis requiring hearing aids, precocious puberty, on physical examination with facial dysmorphias given by prominent broad forehead, mild blue tint sclerae, irregular teeth with dentin wear, ligament hyperlaxity, no cognitive-behavioral compromise and no other alterations. Within the paraclinical approach with hypercalcemia, hypercalciuria and hyperphosphatemia, other paraclinical evaluation of bone metabolism in normal reference range, within the imaging studies with total spine radiography with mild lumbar dextro convex dextro scoliosis, and anterolisthesis of L5 over S1, and radiographs showing areas of consolidation by fractures previously described, renal and urinary tract ultrasound with mild bilateral ectasia and 15% residue in bladder emptying. The patient was the product of non-consanguineous parents, with no known family history of genetic connective tissue diseases. Due to the clinical history and physical examination, a molecular sequencing + CNVs (Copy Number Variations) study was requested by NGS (Next-Generation Sequencing) for genes related to collagenopathies. **Results:** Molecular study sequencing + CNVs by NGS for genes related to collagenopathies in which a heterozygous pathogenic variant c.3652G>A; p.Ala1218Thr in the COL1A1 gene is detected, which supports the diagnosis of osteogenesis imperfecta type I of autosomal dominant inheritance.

Corresponding author

Jhonatan Alzate Valencia

Email

jhonatan-alzatev@unilibre.edu.co

Keywords

Osteogenesis imperfecta, collagen type I, de novo variant, orphan disease, precession medicine.

Bioethical aspects

The author(s) declare that they have no competing interests, and have obtained informed consent from the patients. This work was approved by the institutional ethics committee.

Funding

The authors declare that they have not received external funding associated with this work.

Licence and Distribution

Published by Infomedic International under the Creative Commons Attribution 4.0 International License (CC-BY-NC).

DOI

10.37980/im.journal.ggcl.20242363

Conclusion: Osteogenesis imperfecta is a disease of genetic origin, rare, reported for many years, with phenotypic, endotypic and genotypic variability, so that undertake early identification strategies impact on the natural history of the disease contributing to the development of preventive medicine, personalized, predictive, accurate, participatory in order to be performed at the population level; through this case it is intended to raise awareness of the importance of early diagnosis and genotype-phenotype correlation that allows to establish targeted actions, to know specific pathophysiological mechanisms, to sub-classify collagenopathies, contributing to the increase of medical-scientific knowledge about the expression of the disease and the tendency to hyper-personalization.

INTRODUCTION

Osteogenesis imperfecta (OI), known as a disorder characterized by bone weakness and high risk of fractures, of genetic origin, is known as brittle bone disease [1]. It has been found that this pathology generates a high burden of morbidity and mortality, associated with the presence of pain and functional limitations for those who have it [1]. The fragility present in the bones has been described by structural variants of the Collagen 1 genes (COL1A1 and COL1A2), among others [1,2]. According to population studies, an estimated prevalence between 1/15,000 and 1/20,000 cases in live births is known, being classified as an orphan disease [3,4]. Throughout history, different types of classifications have been presented since the initial classification in 1979 by Sillence, to the clinical classification of 2014, and through classifications based on findings of genetic variants [1,2,16].

According to these classifications, OI can have its etiology in different genetic variants and not only alterations of the COL1A1 collagen gene, however, it is the gene with the greatest implication in this pathology. Any alteration of the long arm of chromosomes 7 and 17 can cause a alteration in the amount of pro-collagen synthesized, with approximately up to half of type I collagen being affected in quantity or quality. So far, more than 150 variants related to this gene have been reported, which can result in OI [6,7]. The main characteristic is the presence of numerous fractures or bone deformities, due to the great fragility, however, there is a great phenotypic heterogeneity [8]. There are severe presentations related to congenital conditions, which include fractures in utero and during childbirth, with serious complications at the brain and lung levels, fatal.

Other cases of mild to moderate presentation, with fewer fractures, more bone deformity, and some other complications. That is why today the clinical classification is based on severity and genetics based on molecular levels [5,7,8]. They also present characteristic facial features, with a wide and prominent forehead with large temporal and parietal bones, blue sclerae due to intraocular pigments, muscular atrophy due to the number of fractures, thin skin, and capillary fragility. 80-90% of patients have scoliosis and basilar impression can be found in up to 2% of patients in

the cervical spine [1,5,6]. Osteogenesis Imperfecta type I is described as the most common presentation, affecting up to two-thirds of the diagnosed population.

It is generally associated with milder severity, with some of the clinical characteristics such as bone fragility, blue sclerae, conductive hearing loss, and minimal bone deformity [1,8]. Timely diagnosis of the disease allows for the initiation of therapeutic alternatives that help reduce the burden of the disease, such as the number of fractures, pain management, and multidisciplinary treatment with growth hormone, anti-resorptive bone therapy, anabolic therapies, or even anti-TGF β antibodies [3]. Numerous research studies on new treatment strategies have been developed with the aim of inhibiting excessive bone resorption or increasing bone formation [8-10].

Among the most promising treatments are denosumab, a monoclonal antibody that acts against the activator of the NF-kappa B ligand receptor, a key cytokine of osteoclasts; and monoclonal antibodies against anti-sclerostin proteins, and dickkopf-1 endogenous inhibitor of bone formation, however these are still in the study phases [11,12]. Additionally, there are studies that report autologous bone marrow transplantation, however, without conclusive results yet. Therefore, a multidisciplinary approach with an emphasis on physical rehabilitation is still being considered to improve motor activity, surgery if needed to correct deformities. Additionally, pharmacological treatment and genetic counseling [11,12] are recommended.

The objective of this clinical case is to raise awareness among healthcare personnel about the importance of timely recognition of this pathology and the impact achieved by making an early and accurate diagnosis of the condition. Despite the phenotypic variability found in this disease, it is of great importance to carry out genetic studies and counseling as they can present as de novo variants, which may appear for the first time in a family, not inherited, secondary to changes in a germ cell (ovum or sperm) or in the zygote; and their identification will allow to start targeted therapeutic alternatives, in order to reduce the burden of morbidity and mortality of the patients who

suffer from it. And provide follow-up guidelines, prognosis, moving closer to personalized and precision medicine.

CLINICAL CASE

Eight-year-old female patient with a clinical history of fractures secondary to mild traumas since the age of 4 years old. The first fracture was in the left foot metatarsal with the mechanism of trauma being a jump from her own height, the second fracture occurred at 6 years old in the right hand finger after falling off a bike, and at 7 years old she fractured her right foot after falling from her own height. Other diagnoses include bilateral otosclerosis requiring hearing aids and early puberty. She is the child of non-consanguineous parents, with no known family history of genetic connective tissue diseases.

During the physical examination, the patient presented with facial dysmorphism associated with a wide, prominent forehead, slightly blue-tinted sclera, irregular teeth with dentin wear, bilateral hearing aids, described scar from fractures, breast bud present, dark and sparse hair without signs of estrogenization, ligamentous hyperlaxity, normal behavior and cognition, with no other alterations on physical examination.

In terms of paraclinical approach, the patient had a report of urinalysis showing urinary sediment calcium +++ and phosphorus of 5.87 mg/dL (elevated), and calcium of 10.56 mg/dL (elevated), with the rest of the paraclinical tests showing normal bone metabolism.

He had imaging studies of total spinal column x-ray with mild dextroconvex lumbar scoliosis, and anterolisthesis of L5 over S1, simple skull x-ray without fractures or alterations, and long bone x-rays - complete series showing previously described fractures, with renal and urinary tract ultrasound revealing mild bilateral ectasia and a 15% residual in bladder emptying.

Due to clinical history and physical examination with suspicion of Collagen Tissue Disease - Collagenopathy, a molecular study (sequencing + CNVs through NGS) was requested for genes related to this medical condition.

RESULTS

Molecular study sequencing + CNVs through NGS for genes related to collagenopathies in which a heterozygous pathogenic variant is detected in the COL1A1 gene (See Table 1).

This variant generates the change of a guanine to an adenine at position 3,652 of the cDNA, in exon 48 of the gene (c.3652G>A), and in the protein it produces the erroneous change of an alanine to a threonine at amino acid 1,218 (p.Ala1218Thr), an evolutionarily conserved amino acid. Another nucleotide change has been previously documented at this position that has been classified as deleterious (chr17:48,264,163 C => G (Ala1218Pro)), which supports the clinical suspicion of hereditary collagenopathy.

Deleterious variants in the COL1A1 gene are associated with autosomal dominant type I osteogenesis imperfecta (see details in supplementary table).

DISCUSSION

In Osteogenesis Imperfecta, there is a variant in the COL1A1 or COL1A2 genes in up to 90% of cases. This affects type I collagen fibers, which play a role in bone strength [1,15].

The Sillence classification [16], which was the first classification in 1976, did not take into account variations in genes and only had some classifications based on clinical criteria. Nowadays, a classification of the disease is carried out both clinically and genetically. In Table 2, the clinical characteristics specific to each type of osteogenesis are observed [11,12,18].

The case described corresponds to a patient with clinical and imaging suspicion of collagenopathy, with molecular confirmation of mild phenotype type I osteogenesis imperfecta, with clinical findings that allow the establishment of said genotype-phenotype correlation, frequent fractures, hearing loss, and blue sclerae. Although otosclerosis is not one of the most frequent findings in the population with osteogenesis imperfecta, there is a study reporting a family with several members affected by both disorders [14].

Table 1. Molecular findings of the patient

Gene	Variant	Zygosity	Meaning Clinical	Illness Associated	Type of Inheritance
COL1A1	c.3652G>A (p.Ala1218Thr)	Heterocigosis	Pathogenic	Osteogenesis Type I imperfecta	Autosomal Dominant

Source: Own elaboration

Table 2. Clinical characteristics according to type of Osteogenesis.

Type I	Lightweight Form Bone fragility Blue Sclera Frequent hearing loss Aortic regurgitation	Type VIII	Similar type I, II or III but with White Sclera Rhizomelia Short stature
Type II	Neonatal Onset Lethal (pulmonary hypoplasia, CNS malformations, hemorrhages) Intrauterine Fractures Dentogenesis imperfecta Blue Sclera	Type IX	Severe deformity Blue Sclera Short stature
Type III	Greater severity in survivors Short stature Severe bone deformity Respiratory failure (pulmonary hypoplasia) Frequent fractures Frequent hearing loss	Type X	Severe deformity Blue Sclera Kidney stones
Type IV	Progressive deformity Short stature Scoliosis Joint laxity Gray or white sclera Dentogenesis imperfecta	Type XI	Severe deformity Muscle contractures
Type V	Moderate Severity Hypertrophic corns at fracture sites Interosseous membrane calcification	Type XII	Frequent fractures Mild deformity Delayed tooth eruption
Type VI	Moderate Severity Rare Defect in bone mineralization (biopsy)	Type XIII	Frequent fractures Hyperlax joints Bone Mass
Type VII	Moderate to severe deformity Shortening of the femur and humerus Blue Sclera Small head round face		

Source: Authors.

According to this study, otosclerosis is present in 12% of Caucasian patients, with a higher prevalence in women, and there are two cases of a family with osteogenesis imperfecta and otosclerosis [1]. However, in our patient, no relatives with other auditory or genetic disorders like osteogenesis imperfecta were found. For the reported inheritance in osteogenesis imperfecta type I, there are findings of autosomal dominant inheritance, mainly in the COL1A1 or COL1A2 genes, but there are also some reported variants in genes CRTAP, PPIB, SP7 [9]. In our patient, molecular study evaluated 68 genes related to collagen disorders (ACTA2, ADAMTS2, ALDH18A1, ATP6V0A2, ATP7A, B3GALT6, B3GAT3, B4GALT7, BGN, C1R, CBS, CHST14, COL11A1, COL12A1, COL1A1, COL1A2, COL2A1, COL3A1, COL4A1, COL5A1, COL5A2, COL5A3, COL6A1, COL6A2, COL6A3, among others) where only this variant was found.

In this case, a change from guanine to adenine was evidenced in the COL1A1 gene at position 3652 of the cDNA, in exon 48 of the gene (c. 3652G>A), which at the protein level produces a change from alanine to threonine at amino acid 1218 (p. Ala1218Thr). As shown in Figure 1 [21], different systems are used to observe the changes presented in the gene and the possible products of these variants. Additionally, a search for this variant was conducted in artificial intelligence assistants. According to the first genetic variant assistant from GenAI, VarChat [22]: the genomic variant c. 3652G>A is a single nucleotide substitution that occurs in the COL1A1 gene, where guanine (G) is replaced by adenine (A) at position 3652 of the DNA nucleotide.

The change results in the alteration of the codon for alanine (Ala) at position 1218 in the protein sequence to a codon for threonine (Thr), referred to as p. Ala1218Thr. The variant is cataloged in public databases with the reference SNP ID rs72656337 (See Figures 1 and 2). Genetic function: the COL1A1 gene encodes the pro-alpha1(I) chain of collagen type I, which is an important structural component of the extracellular matrix in various connective tissues throughout the body. Type I collagen is a fibrillar collagen, formed by the association of two pro-alpha1(I) chains and one pro-alpha2(I) chain, creating a triple helix structure. This protein plays a key role in the strength and elasticity of connective tissues, including bones, skin, and tendons.

Clinical significance

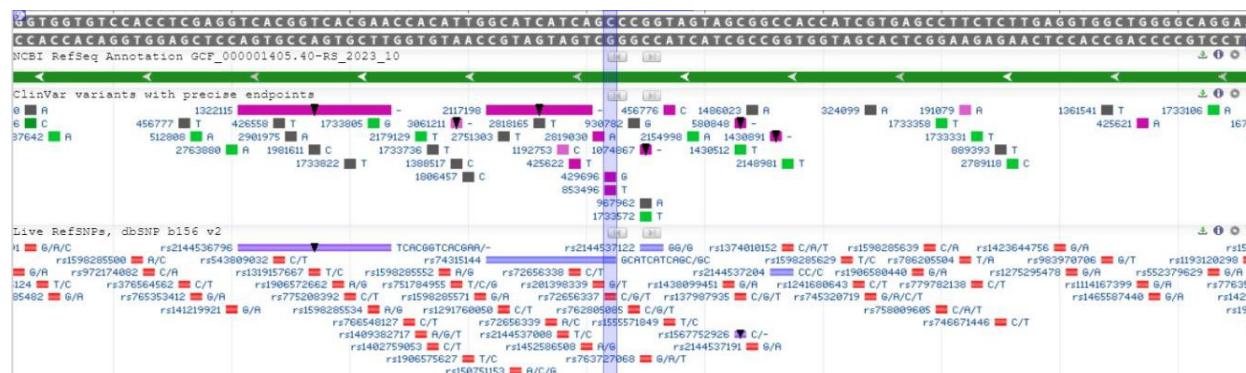
The variant rs72656337 has been reported in ClinVar with a total of 1 pathogenic presentation.

The substitution at position 1218 is located within one of the collagen triple helix domains, which are crucial for the formation and stability of collagen fibrils.

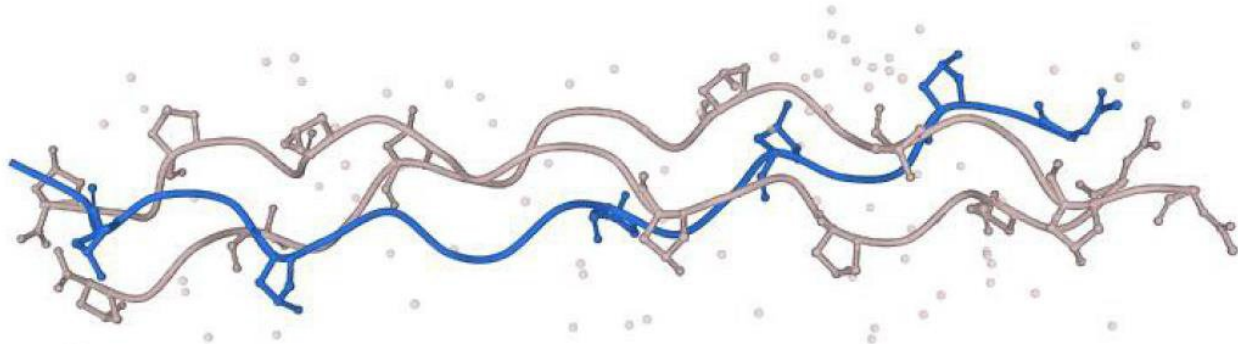
Substitution of the non-polar alanine residue with the polar threonine residue may alter the helical structure, which could lead to changes in the formation and function of the fibrils. These structural changes may have important implications for the mechanical properties of connective tissues.

Pathogenic variants in the COL1A1 gene are known to be associated with various connective tissue disorders, in-

Figure 1. Genomic regions, transcripts and products.



Variante rs72656337, Genoma GRCh38.p15 (NC_000017.11). Tomada de: <https://www.ncbi.nlm.nih.gov/snp/rs72656337>

Figure 2. Representative structure of UniProt 3D AlphaFold of COL1A1.

Note: COL1A1 UniProt 3D 02452 gene. Source: URL: <https://www.ebi.ac.uk/pdbe-kb/proteins/P02452>.

cluding osteogenesis imperfecta (OI), a group of genetic disorders characterized by bone fragility, low bone mass, and susceptibility to fractures. The clinical presentation of OI can vary widely, from mild to severe, depending on the specific nature of the collagen defect.

In conclusion, the c. 3652G>A, p.Ala1218Thr variant in the COL1A1 gene is a nonsense change that has been associated with pathogenic outcomes in the context of connective tissue disorders. The clinical importance reported in ClinVar supports the potential impact of this variant on protein function and disease manifestation. Treatment should be multidisciplinary and based on the patient's clinical condition. Recent studies have shown the use of Setrusumab as a biological agent in managing bone calcium fixation. However, these studies have only been conducted in adults [19]. There is only one study in children comparing the use of Setrusumab and bisphosphonates, but this study is still in phase 3, with further studies needed on the prevalence of this type of osteogenesis and pharmacogenomics [20].

CONCLUSION

Osteogenesis imperfecta is a rare genetic disease with phenotypic, endotypic, and genotypic variability, which is why undertaking strategies for early identification impacts the natural history of the disease, contributing to the development of preventive, personalized, predictive, precise, participatory medicine in order to be carried out at a population level. The different databases, different high-performance bioinformatics algorithms, individual predictors, and MetaScore, together with knowledge about functionality, biological bases, genomic and molecular annotation data,

protein structure and function, functional studies, use of artificial intelligence tools, and in accordance with Richards, et al.

Standards and guidelines for the interpretation of sequence variants [23], by the American College of Medical Genetics and Genomics, Association for Molecular Pathology, ClinGen, classify this variant as likely pathogenic. Using the artificial intelligence genomic platform Mastermind, it is classified as PM2, PM1, PP2, PP1M, PPC, PPC Het with NM_000088.4:c.3652G>A (p.Ala1218Thr) Pathogenic, which establishes a genotype/endotype/phenotype correlation. Through this case, the importance of a diagnosis and genotype-phenotype correlation that allows for targeted actions to be taken, understanding specific pathophysiological mechanisms that allow for the subclassification of collagenopathies, contributing to the increase in medical-scientific knowledge about disease expression and the trend towards hyper-personalization, is sought to be raised awareness.

ABOUT THE AUTHORS

Jhonatan Alzate-Valencia

Physician, Resident Specialization in Pediatrics, Research Group in Pediatrics (GRINPED), Neurogenetics and metabolic diseases research line.

Lina-Johanna Moreno-Giraldo

Physician, Specialist in Pediatrics, Master and PhD in Medical Genetics, Research Group in Pediatrics (GRINPED), Neurogenetics and Metabolic Diseases Research Line. Professor, Department of Clinical Postgraduate Studies.

REFERENCES

- [1] Ibáñez, A., & Hodgson, F. (2021). OSTEOGÉNESIS IMPERFECTA. *Revista Médica Clínica las Condes*, 32(3), 311-318. <https://doi.org/10.1016/j.rmcl.2020.09.004>
- [2] Torrent, B. (2020). Osteogénesis imperfecta (2.a ed.). <https://www.aeped.es/protocolos/ISSN 2171-8172>
- [3] Rossi, V., Lee, B., & Marom, R. (2019). Osteogenesis imperfecta: advancements in genetics and treatment. *Current Opinion In Pediatrics*, 31(6), 708-715. <https://doi.org/10.1097/mop.0000000000000813>
- [4] Deguchi, M., Tsuji, S., Katsura, D., Kasahara, K., Kimura, F., & Murakami, T. (2021). Current Overview of Osteogenesis Imperfecta. *Medicina-lithuania*, 57(5), 464. <https://doi.org/10.3390/medicina57050464>
- [5] Zhytnik, L., Maasalu, K., Reimand, T., Duy, B. H., Kõks, S., & Märtson, A. (2020). Inter- and intrafamilial phenotypic variability in individuals with collagen-related Osteogenesis Imperfecta. *Clinical And Translational Science*, 13(5), 960-971. <https://doi.org/10.1111/cts.12783>
- [6] Maioli, M., Gnoli, M., Boarini, M., Tremosini, M., Zambrano, A., Pedrini, E., Mordenti, M., Corsini, S., D'Eufemia, P., Versacci, P., Celli, M., & Sangiorgi, L. (2019). Genotype– phenotype correlation study in 364 osteogenesis imperfecta Italian patients. *European Journal Of Human Genetics*, 27(7), 1090-1100. <https://doi.org/10.1038/s41431-019-0373-x>
- [7] Salacińska, K., Michałus, I., Pinkier, I., Rutkowska, L., Chlebna-Sokół, D., Jakubowska- Pietkiewicz, E., Kępczyński, Ł., Salachna, D., & Gach, A. (2022). The first glycine-to-tryptophan substitution in the COL1A1 gene identified in a patient with progressively-deforming Osteogenesis imperfecta. *Molecular Genetics & Genomic Medicine*, 10(8). <https://doi.org/10.1002/mgg3.1996>
- [8] Yang, K., Liu, Y., Wu, J., Zhang, J., Hu, H., Yan, Y., Chen, W., Yang, S., Sun, L., Sun, Y., Wu, Q., & Yin, C. (2022). Prenatal Cases Reflect the Complexity of the COL1A1/2 Associated Osteogenesis Imperfecta. *Genes*, 13(9), 1578. <https://doi.org/10.3390/genes13091578>
- [9] Jung, H., Rim, Y. A., Park, N., Nam, Y., & Ju, J. H. (2021). Restoration of Osteogenesis by CRISPR/ Cas9 Genome Editing of the Mutated COL1A1 Gene in Osteogenesis Imperfecta. *Journal Of Clinical Medicine*, 10(14), 3141. <https://doi.org/10.3390/jcm10143141>
- [10] Yang, L., Liu, B., Dong, X., Wu, J., Sun, C., Li, X., Cheng, R., Wu, B., Wang, H., Tong, S., Wang, D., & Luo, F. (2022). Clinical severity prediction in children with osteogenesis imperfecta caused by COL1A1/2 defects. *Osteoporosis International*, 33(6), 1373-1384. <https://doi.org/10.1007/s00198-021-06263-0>
- [11] Dirani, M., Cuenca, V. D., & Romero, V. (2022). COL1A1 novel splice variant in osteogenesis imperfecta and splicing variants review: A case report. *Frontiers In Surgery*, 9. <https://doi.org/10.3389/fsurg.2022.986372>
- [12] Zhuang J, Chen C, Chen Y, Luo Q, Wang Y, Jiang Y, Zeng S, Xie Y and Chen D (2022) Identification of a Rare Variant of c.1777G>A (p.G593S) in the COL1A1 Gene as the Etiology of Recurrent Osteogenesis Imperfecta by Whole-Exome Sequencing. *Front. Pediatr.* 10:816090. doi:10.3389/fped.2022.816090
- [13] Zhytnik L, Maasalu K, Pashenko A, Khmyzov S, Reimann E, Prans E, Kõks S and Märtson A (2019) COL1A1/2 Pathogenic Variants and Phenotype Characteristics in Ukrainian Osteogenesis Imperfecta Patients. *Front. Genet.* 10:722. doi:10.3389/fgene.2019.00722
- [14] Lee, H. N., Jeon, H. J., & Seo, Y. J. (2021b). Familial Otosclerosis Associated with Osteogenesis Imperfecta: A Case Report. *Journal Of Audiology & Otology*, 25(4), 230- 234. <https://doi.org/10.7874/jao.2021.00122>
- [15] Mejia de Beldjenna, L., Lambraño, A., & Mejia, V. (2023). Osteogenesis Imperfecta due to Frameshift Mutation of the Col1a1 Gene. *EC ORTHOPAEDICS*, 14(3).

- [16] Sillence, D., Senn, A., & Danks, D. M. (1979). Genetic heterogeneity in osteogenesis imperfecta. *Journal Of Medical Genetics*, 16(2), 101-116. <https://doi.org/10.1136/jmg.16.2.101>
- [17] Franzone, J. M., Shah, S. A., Wallace, M., & Kruse, R. W. (2019). Osteogenesis imperfecta. *Orthopedic Clinics Of North America*, 50(2), 193-209. <https://doi.org/10.1016/j.ocl.2018.10.003>
- [18] Chimbo, P. F. T., Toala, O. J. R., Andrade, E. F., & Pinto, F. F. J. (2019). Osteogénesis imperfecta: revisión de la literatura actual. *ECUATORIANA DE PEDIATRÍA*, 20(1), 4-9]. <https://pesquisa.bvsalud.org/portal/resource/pt/biblio-1010308>
- [19] Liu, W., Nicol, L., & Orwoll, E. (2024). Current and Developing Pharmacologic Agents for Improving Skeletal Health in Adults with Osteogenesis Imperfecta. *Calcified Tissue International*. <https://doi.org/10.1007/s00223-024-01188-2>
- [20] Setrusumab vs bisphosphonates in children from 2 to < 7 years old with osteogenesis imperfecta (OI). (s. f.). Yale Medicine. <https://www.yalemedicine.org/clinical-trials/oi-study-age-2-to-7-years>
- [21] dbSNP, rs72656337 RefSNP Report - NCBI. (2022, 21 septiembre). dbSNP. <https://www.ncbi.nlm.nih.gov/snp/rs72656337>
- [22] De Paoli, F., Berardelli, S., Limongelli, I., Rizzo, E., & Zucca, S. (2024). VarChat: the generative AI assistant for the interpretation of human genomic variations. *Bioinformatics*. <https://doi.org/10.1093/bioinformatics/btae183>.
- [23] Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, Grody WW, Hegde M, Lyon E, Spector E, Voelkerding K, Rehm HL; Comité de Garantía de Calidad del Laboratorio ACMG. Estándares y directrices para la interpretación de variantes de secuencia: una recomendación de consenso conjunta del Colegio Americano de Genética Médica y Genómica y la Asociación de Patología Molecular. *Genet Med*. 2015 Mayo; 17(5):405-24. doi: 10.1038/gim.2015.30. Epub 5 de marzo de 2015. PMID: 25741868; PMCID: PMC4544753.